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Abstract: AIMS OF THE STUDY Combination antiretroviral therapy (cART) has reduced mother-to-child transmissions (MTCT) and improved the prognosis of HIV-infected newborns. However, drug resistance mutations (DRM) in HIV-infected children, either transmitted by MTCT (HIV-tDRM) or selected by suboptimal adherence and drug levels (HIV-sDRM), remain a concern. We sought to determine the rate of HIV-tDRM and HIV-sDRM in MTCT pairs in Switzerland. **METHODS** We performed a retrospective analysis of prospectively collected clinical data and available stored samples from MTCT pairs participating in the Swiss Mother-Child HIV (MoCHIV) cohort. **RESULTS** We identified 22 HIV-infected mother-child pairs with delivery between 1989 and 2009 who had 15 years of follow-up (33% white ethnicity). Twenty-one women (96%) were treatment-naïve before pregnancy, 8 (36%) had an unknown HIV status and delivered vaginally, 2 were diagnosed but not treated, and 11 (50%) received antiretrovirals during pregnancy or at delivery, of whom only 6 cases (27%) had cART. HIV subtypes were concordant in all mother-child pairs (subtype B 13/22 [59%]). Using stored plasma (n = 66) and mononuclear cell (n = 43) samples from the children, HIV-tDRM (M184V) was identified in 1 of 22 (4.5%) mothers (1/11 treated, 9%) and was followed by HIV-sDRM at 10 months of age. HIV-sDRM (M184V 23%; K103N 4.5%; D67N 13.6%) occurred in 16/22 (73%) after 4 years, half of whom were treatment naïve. HIV-sDRM were associated with a lower CD4 T-cell nadir (p < 0.05) and tended to have higher viral loads and more frequent cART changes. **CONCLUSIONS** HIV-tDRM were low in this Swiss MoCHIV cohort, making them a minor yet preventable complication of prenatal HIV care, whereas HIV-sDRM are a significant challenge in paediatric HIV care.

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The rate of mother-to-child transmission of antiretroviral drug-resistant HIV strains is low in the Swiss Mother and Child HIV Cohort Study

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Summary

AIMS OF THE STUDY: Combination antiretroviral therapy (cART) has reduced mother-to-child transmissions (MTCT) and improved the prognosis of HIV-infected newborns. However, drug resistance mutations (DRM) in HIV-infected children, either transmitted by MTCT (HIV-tDRM) or selected by suboptimal adherence and drug levels (HIV-sDRM), remain a concern. We sought to determine the rate of HIV-tDRM and HIV-sDRM in MTCT pairs in Switzerland.

METHODS: We performed a retrospective analysis of prospectively collected clinical data and available stored samples from MTCT pairs participating in the Swiss Mother-Child HIV (MoCHIV) cohort.

RESULTS: We identified 22 HIV-infected mother-child pairs with delivery between 1989 and 2009 who had 15 years of follow-up (33% white ethnicity). Twenty-one women (96%) were treatment-naïve before pregnancy, 8 (36%) had an unknown HIV status and delivered vaginally, 2 were diagnosed but not treated, and 11 (50%) received antiretrovirals during pregnancy or at delivery, of whom only 6 cases (27%) had cART. HIV subtypes were concordant in all mother-child pairs (subtype B 13/22 [59%]). Using stored plasma (n = 66) and mononuclear cell (n =

43) samples from the children, HIV-tDRM (M184V) was identified in 1 of 22 (4.5%) mothers (1/11 treated, 9%) and was followed by HIV-sDRM at 10 months of age. HIV-sDRM (M184V 23%; K103N 4.5%; D67N 13.6%) occurred in 16/22 (73%) after 4 years, half of whom were treatment naïve. HIV-sDRM were associated with a lower CD4 T-cell nadir (p < 0.05) and tended to have higher viral loads and more frequent cART changes.

CONCLUSIONS: HIV-tDRM were low in this Swiss MoCHIV cohort, making them a minor yet preventable complication of prenatal HIV care, whereas HIV-sDRM are a significant challenge in paediatric HIV care.

Keywords: mother-to-child transmission, drug resistance mutation, HIV-tDRM, HIV-sDRM, combination antiretroviral therapy

Introduction

Mother-to-child transmission (MTCT) accounts for more than 90% of HIV infections during childhood according to UNAIDS reports [1, 2]. Missed maternal diagnosis and non-treatment are the main causes of MTCT. These are preventable by a combination of measures consisting of early HIV testing for all women as part of routine prenatal care, immediate combined antiretroviral treatment (cART) and regular monitoring throughout pregnancy. Complete

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HIV load suppression by the third trimester, and specifically at the time of delivery, reduces the risk of transmission to almost zero. As outlined by the WHO (Prevention of MTCT – PMTCT) and European guidelines [3, 4], treatment-naïve women should start cART immediately, with an integrase strand transfer inhibitor (INSTI)-containing regimen being the preferred treatment. Pregnant women already under suppressive cART should continue, but contraindicated drugs such as didanosine, stavudine or triple nucleoside reverse transcriptase inhibitor (NRTI) combinations should be replaced. The latest data from the IMPAACT (International Maternal Pediatric Adolescent AIDS Clinical Trials Network) study suggest that exposure of mothers to dolutegravir (DTG) at the time of conception could be associated with a higher risk of neural tube defects. Therefore, the Swiss HIV Cohort Study (SHCS) advises against prescribing DTG for women who are trying to become pregnant [5].

In case of incomplete suppression (HIV-VL >50 copies (cp)/ml) at 34–36 weeks of gestation, intravenous zidovudine administration should be considered during labour and Caesarean section. In the case of ongoing maternal viral replication, international guidelines recommend starting cART in high-risk newborns as soon as possible and advocate lifelong continuation in the case of confirmed MTCT [4, 6, 7]. Recent studies indicate that starting cART at birth is associated with improved long-term outcomes for HIV-infected children [8, 9]. Accordingly, morbidity and mortality in HIV-infected children have significantly decreased in the recent decades [10].

HIV-infected children in Western Europe and North America benefit from easy access to specialised medical care which provides laboratory monitoring and cART [11]. Moreover, novel drugs and combinations have facilitated cART for pregnant women and children alike, allowing the goal of long-term HIV suppression with outstanding benefits for health at the individual, societal, and epidemiological levels to be reached.

Nevertheless, virological failure remains a key challenge in the management of HIV-infected children. The rates of virological failure range from 12% to 32% in high and limited-resource settings, respectively. Rates peak during infancy, again during adolescence, and then when transitioning to adult medical care [12–14]. Although dosing of cART in children is sometimes difficult because a suitable drug formulation (e.g. suspension) is not available, virological failure reflects not only a lack of access to care and cART or toxicity concerns, but often also adherence issues of both mothers and children. Thus, a main challenge in HIV-infected children is the emergence of HIV variants with drug resistance mutations (DRM), either selected by suboptimal adherence and insufficient drug combinations and levels (HIV-sDRM), or originating from MTCT (HIV-tDRM). For the reasons pointed out earlier, the risk of HIV-sDRM increases during the HIV-infected child's lifetime [14]. Conversely, MTCT may involve HIV-tDRM [15], which may also impact cART efficacy and favour the emergence of HIV-sDRM. Here, we report the results of a retrospective analysis of data and blood samples collected prospectively from MTCT pairs participating in the Swiss Mother and Child HIV Cohort Study (MoCHIV) in order

to evaluate the rate of transmitted and selected HIV drug resistance in Switzerland.

Materials and methods

Study design

Data and blood samples were collected prospectively by MoCHIV, in which 277 HIV-infected children have been enrolled since 1989. These children were followed in clinics in Basel, Bern, Genève, St Gallen, Zurich and Ticino [16]. MoCHIV is approved by the Swiss Federal Office of Public Health and by the institutional review boards of the participating centres. All women gave their written informed consent to participate in the study, including the retrospective analysis of their and their children's anonymised data and stored material. In June 2017, 274

children had been registered in Switzerland as being born to HIV-infected mothers (see [MoCHIV on SHCS website](#)). Of these, 1559 children were diagnosed as being not infected with HIV-1, 145 children had an undefined HIV-status, and 277 children were HIV-1 infected and participated in the Swiss MoCHIV. Of the HIV-positive children, 72 were lost from follow-up or are being treated by a non-cohort physician, 62 have died, and 59 are currently under follow-up in MoCHIV.

For inclusion in this virological DRM study, we identified all MTCT pairs enrolled in the MoCHIV cohort from whom samples were available from the mothers at the time of pregnancy and/or within one year postnatally, and from the corresponding MoCHIV follow-up visits of the children. We analysed demographic, clinical and laboratory parameters including plasma HIV load, CD4 T-cell counts, cART, adherence and clinical conditions, as well as prospectively cryopreserved plasma and whole-blood samples. All available genotype and drug resistance profiles previously performed were included in the analysis. In addition, HIV-DRM were identified by HIV genotyping and resistance testing of the samples from mothers and children at the time of, or shortly after, birth in the Laboratory for Genotyping and Resistance Testing (formerly Division of Infection Diagnostics, Department Biomedicine, University of Basel, accreditation STS219; from 1 Jan 2019: Clinical Virology, Laboratory Medicine, University Hospital Basel, STS0568) to identify tDRM. HIV-tDRM were defined as DRM detected post-natally in RNA or proviral DNA samples that had been obtained before any antiretroviral treatment was administered and which matched the mother's viral genotype. HIV-sDRM were defined as new DRM arising in children with virological failure.

HIV genotyping and resistance testing

The standard of care of HIV genotyping and resistance testing in our institution is Sanger sequencing of amplicons generated from the HIV protease, reverse transcriptase and, since 2012, the integrase genes. This analysis was performed on both plasma and/or peripheral blood mononuclear cell (PBMC) samples by generating amplicons using the Abbott ViroSeq HIV-1 Genotyping kit (Abbott, IL, USA) or in-house methods validated and accredited in the Division of Infection Diagnostics, Department Biomedicine, University of Basel (STS219). Briefly, following nested PCR and quality control by agarose gel elec-

trophoresis, the amplicons were purified with illustra ExoProStar 1-Step (GE Healthcare, England). The sequencing was performed using the BigDye Terminator v3.1 cycle sequencing kit (Thermo Fisher Scientific, MA, USA), purification with Sephadex G-50 (GE Healthcare, England) and capillary electrophoresis on a 3500 Genetic Analyzer (Thermo Fisher Scientific, MA, USA). The sequences were analysed using SmartGene (Lausanne, Switzerland) [17] and the results interpreted using the [HIV Drug Resistance Database of Stanford University](#) (CA, USA). As an accredited regional HIV reference laboratory, the Laboratory for Genotyping and Resistance Testing participates in an external quality assurance program using the French ANRS.

Additionally, deep sequencing using the Illumina MiniSeq platform (CA, USA) was performed on 10 samples from three mother and child pairs. HIV protease, reverse transcriptase and integrase were first amplified using nested PCR primers, followed by library preparation using the Illumina Nextera XT kit (CA, USA). Paired-end sequencing was performed using the Illumina MiniSeq MidOutput Kit and a read length of 150bp. Reports were generated by DeepChek-HIV (ABL, Luxembourg) [18]. Proviral load was determined using DNA extracted from the collected PBMCs using a quantitative, real-time PCR assay with normalisation to 150,000 cells using aspartoacylase, a human diploid housekeeping gene routinely used for this purpose [19].

Data analysis

Patient demographics and laboratory parameters were analysed using descriptive statistics. Data from each individual child were described longitudinally over time. Demographic characteristics of mother-child pairs are presented as proportions (percentages) or as medians (25th, 75th, range), using box and whisker plots where indicated. Categorical variables were compared using the unpaired t-test and Fisher's exact test, whereas continuous variables were analysed using the Wilcoxon rank-sum test. Two-sided p-values of <0.05 were considered statistically significant. Analyses were performed using the statistical software package SPSS version 24 (IBM® SPSS Statistics

24.0.0.1) and Excel version 15.37 (Microsoft® Excel 2016 for Mac).

Results

We identified 22 MTCT pairs which fulfilled the enrolment criteria in the Swiss MoCHIV study. The children were born between 1989 and 2009, thereby covering 20 years of different treatments and antiretroviral drug availabilities. Nineteen children are still enrolled in this national cohort and three have moved to the adult Swiss HIV Cohort Study (SHCS). Median gestational age at delivery was 38 weeks (25th percentile 28, 75th percentile 39, range 27–42 weeks) and 27% of the children were born prematurely (<37 weeks) ([table 1](#)). Eight of the 22 children (36%) were delivered vaginally. All but one of the mothers (21/22, 95%) reported as treatment-naïve before pregnancy. During pregnancy, only 11 (50%) mothers received any kind of antiretroviral therapy (zidovudine in five cases, [table 1](#)), and cART was administered to only 6 (27%) mothers. Plasma HIV load at the time of delivery was only available for 10 mothers, and none of them were suppressed to <50 cp/ml (median 71,249 cp/ml; [table 1](#)).

The demographics of the HIV-infected children are summarised in [table 2](#). To the best of our knowledge, there were no siblings in this analysis, but this was not due to explicit exclusion. Five (23%) children had detectable plasma HIV loads at birth or within the first month, suggesting *in utero* transmission (median HIV load 13,845 cp/ml; range 6880–2,531,122). In another 14 children, the first plasma HIV loads were detected within the first nine months of life. Only one child had undetectable HIV loads at six months of age, whereas the other children had median plasma HIV loads of 200,000 cp/ml (IQR 16,274–765,085; range 748–2,531,122). During follow-up visits, the nadir CD4 T-cell count varied widely, between 14 and 1,282/μl (median 280; IQR 146–627). Peak HIV loads at the time of diagnosing virological failure ranged from 1160 to 7,400,000 cp/ml (median 424,913; IQR 64,831–958,025). At last follow-up (median 14 years old; IQR 14–18 years; range 6–23 years), 19 of the 22 children were alive and attending regular follow-up (one child died at one year of age, two children were lost to follow-up at

Table 1: Clinical characteristics of mothers in MTCT pairs.

Variables		n (%) or median	25th, 75th percentiles (range)
Gestational age at delivery (weeks)		38	28, 39 (27–42)
Type of delivery	Vaginal	8 (36)	
	Caesarean	11 (50)	
	Unknown	3 (14)	
Maternal complication at delivery	Haemorrhage	2 (9)	
	Infection	4 (18)	
	Other	2 (9)	
ART during pregnancy	Yes	11 (50)	
	Duration ≤1month	8 (73)	
	Duration >1month	3 (27)	
Type of therapy / prophylaxis	cART	6 (27)	
	ZDV	3 (14)	
	ZDV intrapartum	2 (9)	
	None	11 (50)	
CD4 T cell at birth (cells/μl)		308	72, 580 (32–822)
HIV load at birth (copies/ml)		71,249	9840, 132,621 (204–284,000)

cART = combination antiretroviral therapy; ZDV = zidovudine

9 and 10 years old), and 16 of these 19 children were on cART. The CD4 cell counts had a median of 709 cell/mm³ (IQR 555–848; range 240–1,300). Plasma HIV loads were below the limit of detection (<20 cp/ml) in 11/19 children (median 60 cp/ml; IQR 0–77; range 0–10,000).

Antiviral treatment and virological failure

Thirteen (59%) children were born before the year 2000. Individual infant treatment history from the start of ART is summarised in [figure 1](#). All children, except one who died within the first year of life, had a follow-up longer than two years. Among children who received cART, there was an average of five treatment changes (median; IQR 3–8; range 2–16) ([table 2](#)). Virological failure occurred in all children during follow-up (median 2; IQR 2–3.5; range 1–6). Adjusting the observational data for the year of effective follow-up indicated an average of 0.18 virological failure episodes per year and child, i.e., a virological failure occurred once every five years of observation time on average. Approximately half the virological failure events were among those older than eight years of age. In three children, plasma HIV loads were detectable over prolonged periods of more than 10 years.

Genotype and drug resistance profile

A total of 165 genotypic resistance profiles were analysed. One third of them (56/165) had been performed previously during a routine visit. For 33 profiles, only partial sequences were available, and these needed to be repeated. Complete sequences of sufficient quality were available for a total of 109 samples, consisting of 66 plasma- and 43 PBMC-derived sequences. Concordant HIV-1 subtypes were observed in all mother-child pairs, the most prevalent being subtype B in 13/22 (59%) ([fig. 2](#)). Other subtypes and recombinant strains included C in 18% (4/22) of mother-child pairs and AC, A1C, A1, CRF18_cpx and

CRF67_01B in 4.5% (1/22) each. Since protease inhibitors were the first antiretrovirals to be used in cART in Switzerland in 1996 [20], we used HIV-protease sequences as a surrogate of diversity and evolution over time. In the circular phylogram of mother and child viral RNA and proviral DNA sequences ([fig. 2](#)), examples of viral evolution are visualised in the distance between branches, which are still clustered within their respective HIV-1 subtypes. HIV polymorphisms and sequences not conferring intermediate to high-level antiretroviral resistance were detected in all children (not shown). HIV resistance genotypes conferring relevant intermediate to high-level antiretroviral resistance were found in 45% (10/22) of the mothers, affecting susceptibility to protease inhibitors (PI) in seven (70%), to NRTI in one (10%), to non-nucleo(s)tide reverse transcriptase inhibitors (NNRTI) in three (30%), and to INSTI-containing therapy in one (10%).

HIV-tDRM were identified in one of the 22 MTCT pairs (4.5%), showing matched mother- and child-derived HIV sequences (1 of 11 ART-exposed mothers, 9%). The presence of HIV-tDRM was associated with early emergence of HIV-sDRM (0.83 vs 3.5 years after birth), but the small numbers precluded a meaningful statistical analysis.

The emergence of HIV-sDRM was seen in 16/22 (73%) children, at a mean age of 4.3 years (median 2.8; IQR 0.8–5.9; range 0.25–14.8), and 10/22 (45%) had two or more mutations. Thirteen of these 16 (81%) were on treatment at the time of HIV-sDRM emergence. Children with HIV-sDRM showed a trend towards a greater number of treatment changes over the entire follow-up period compared to children without HIV-sDRM (7 vs 4 times, respectively; $p = 0.104$). The CD4 T-cell nadir was lower and associated with higher HIV loads in children with HIV-sDRM ($p < 0.05$ and $p = 0.06$, respectively; [fig. 3](#)).

The most frequent DRM was M184V, detected in 7/22 (32%), while other major NRTI mutations (M41L, L210W,

Table 2: Demographics of the HIV-infected children.

Variables		n (%) or median	25th, 75th percentiles (range)
Total		22 (100)	
Gender (female)		11 (50)	
Ethnicity	White	7 (32)	
	Black	8 (36)	
	Other	7 (32)	
HIV-1 subtype	B	13 (59)	
	Non-B	9 (41)	
Peak HIV load (copies/ml)		424,914	64,831, 958,025 (1160–7,400,000)
Nadir CD4 T cell (cells/ μ l)		280	146,627 (14–1,282)
Antenatal prophylaxis (yes)		8 (36)	
Age of the children at ART start	At birth	10 (45)	
	Not at birth but ≤ 1 year	7 (32)	
	Later >1 year	4 (18)	
	Never	1 (5)	
Initial HIV treatment	ART	11 (50)	
	cART	10 (45)	
	No therapy	1 (5)	
CD4 T-cells at ART start (counts/ μ l)		1,579	379, 2,45 (65–2,649)
HIV load at ART start (copies/ml)		181,256	13,278, 602,277 (0–8,829,994)
ART changes (number)		5	3, 8 (2–16)

ART = antiretroviral therapy; cART = combination antiretroviral therapy

commonly M46I, followed by V82A and I54V. Only one major INSTI DRM (R263K) was detected, in one child in a proviral sequence, while minor INSTI DRM were found in a total of six patients. However, clinical evidence of associated virological failure was lacking, as none of the patients were exposed to this class of drugs. In all but two cases, HIV-DRM emerged after previous exposure ranging

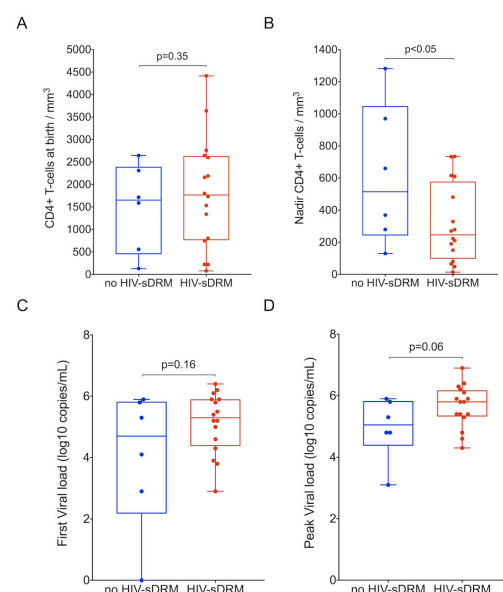
[illegible]

Phylogenetic tree showing the relationships between HIV-1 CRF67_01B and CRF18_01B sequences. The tree is rooted at the bottom and branches upwards. The sequences are grouped into five main clades, each enclosed in a colored oval:

- Subtype A1C** (green oval): Includes sequences such as F01791.16.02.2009 PM, F01801.08.05.1989 PM, and T00791.17.06.1990 PM.
- Subtype A1** (light blue oval): Includes sequences such as Z00791.20.08.2009 PM, Z00791.08.05.2003 PM, and Z00791.10.04.2003 PM.
- Subtype CRF18_01B** (yellow oval): Includes sequences such as H01791.08.10.2003 PM and H01791.01.08.2003 PM.
- Subtype C** (pink oval): Includes sequences such as T00791.12.04.1987 PM, T00791.20.11.1987 PM, and T00791.13.01.1988 PM.
- Subtype B** (large light blue oval): Includes sequences such as H01791.16.08.1988 PM, H01791.17.06.1990 PM, H01791.18.05.1987 PM, H01791.19.05.1987 PM, H01791.20.08.2009 PM, H01791.21.08.2009 PM, H01791.22.08.2009 PM, H01791.23.08.2009 PM, H01791.24.08.2009 PM, H01791.25.08.2009 PM, H01791.26.08.2009 PM, H01791.27.08.2009 PM, H01791.28.08.2009 PM, H01791.29.08.2009 PM, H01791.30.08.2009 PM, H01791.31.08.2009 PM, H01791.32.08.2009 PM, H01791.33.08.2009 PM, H01791.34.08.2009 PM, H01791.35.08.2009 PM, H01791.36.08.2009 PM, H01791.37.08.2009 PM, H01791.38.08.2009 PM, H01791.39.08.2009 PM, H01791.40.08.2009 PM, H01791.41.08.2009 PM, H01791.42.08.2009 PM, H01791.43.08.2009 PM, H01791.44.08.2009 PM, H01791.45.08.2009 PM, H01791.46.08.2009 PM, H01791.47.08.2009 PM, H01791.48.08.2009 PM, H01791.49.08.2009 PM, H01791.50.08.2009 PM, H01791.51.08.2009 PM, H01791.52.08.2009 PM, H01791.53.08.2009 PM, H01791.54.08.2009 PM, H01791.55.08.2009 PM, H01791.56.08.2009 PM, H01791.57.08.2009 PM, H01791.58.08.2009 PM, H01791.59.08.2009 PM, H01791.60.08.2009 PM, H01791.61.08.2009 PM, H01791.62.08.2009 PM, H01791.63.08.2009 PM, H01791.64.08.2009 PM, H01791.65.08.2009 PM, H01791.66.08.2009 PM, H01791.67.08.2009 PM, H01791.68.08.2009 PM, H01791.69.08.2009 PM, H01791.70.08.2009 PM, H01791.71.08.2009 PM, H01791.72.08.2009 PM, H01791.73.08.2009 PM, H01791.74.08.2009 PM, H01791.75.08.2009 PM, H01791.76.08.2009 PM, H01791.77.08.2009 PM, H01791.78.08.2009 PM, H01791.79.08.2009 PM, H01791.80.08.2009 PM, H01791.81.08.2009 PM, H01791.82.08.2009 PM, H01791.83.08.2009 PM, H01791.84.08.2009 PM, H01791.85.08.2009 PM, H01791.86.08.2009 PM, H01791.87.08.2009 PM, H01791.88.08.2009 PM, H01791.89.08.2009 PM, H01791.90.08.2009 PM, H01791.91.08.2009 PM, H01791.92.08.2009 PM, H01791.93.08.2009 PM, H01791.94.08.2009 PM, H01791.95.08.2009 PM, H01791.96.08.2009 PM, H01791.97.08.2009 PM, H01791.98.08.2009 PM, H01791.99.08.2009 PM, H01791.100.08.2009 PM.

from six months to 10 years. DRM were already present at the time of the first genotypic analysis (M184V, T215Y, L210W, M230I) in only two children without medical history of prior ART.

Figure 3: HIV load and CD4 T-cell count in vertically HIV-infected children. A. CD4 T-cell count in MTCT children at birth with and without selected HIV drug resistance mutations (HIV-sDRM). B. Nadir CD4 T-cell count in MTCT children with and without HIV-sDRM. C. First HIV-1 load in MTCT children with and without HIV-sDRM. D. Peak HIV-1 load in MTCT children with and without HIV-sDRM. Box and whiskers plots showing median and 25th and 75th percentiles.



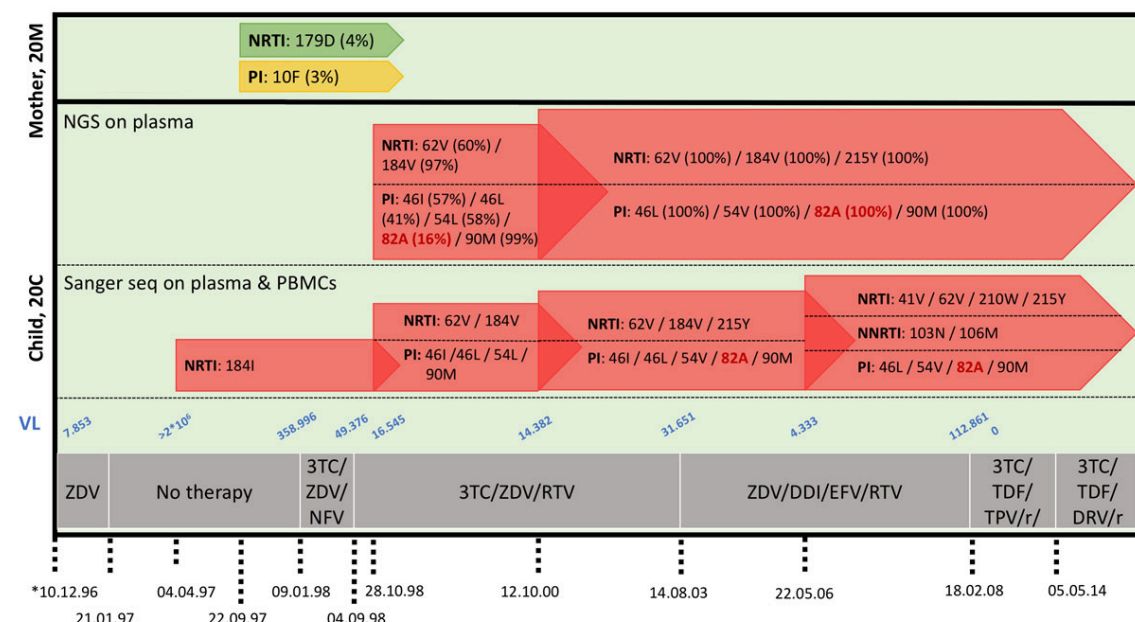
In three MTCT pairs, 10 sample materials from the time of delivery were available in sufficient amounts to complement the conventional Sanger sequencing by next-generation sequencing (NGS). In pair 21, no additional DRM were found. In pair 18, DRM M184I and V, 41L, 65R, 67N, 70R, 74I, 210W and 215Y were detected at frequencies of 53.7% (1,300 reads) and 22.7% (551 reads), 63.7% (557 reads), 1.2% (22 reads), 70.6% (1,282 reads), 1.2% (25 reads), 3% (64 reads), 4.1% (46 reads) and 70% (609 reads), respectively. None of the variants with frequencies of <4% were seen by amplicon-directed Sanger sequencing. In pair 20, NGS identified the PI resistance 82A at a rate of 16.5% (44,670 reads) during the time when the child had persistent high HIV loads having been exposed to this drug class. The PI resistance 82A increased to a detection rate of 99.6% (147,899 reads) two years later with the same method, and at that time was also identified by Sanger sequencing (fig. 4).

Discussion

In this study of the Swiss MoCHIV cohort, we analysed HIV-1 genotype and drug resistance profiles in data and blood samples collected prospectively from 22 HIV infected mother-child pairs. The rate of HIV-tDRM was one out of all 22 MTCT pairs (4.5%) and 1/11 (9%) MTCT pairs where mothers had been previously exposed to antiretrovirals. Although the overall study population was too small for more robust conclusions, other studies have reported rates between 4.9% and 17% [21–23]. The HIV-tDRM detected affected the HIV reverse transcriptase through the M184V mutation.

Conversely, the rate of HIV-sDRM was 16/22 (73%), emphasising that HIV-sDRM represent a significant issue in

Figure 4: Deep sequencing of samples from mother-child pair 20. Comparison of Sanger sequencing and deep sequencing results of HIV-1 RNA from mother-child pair 20. Sampling dates and treatment history are indicated, as well as HIV-1 loads (copies / mL). Deep sequencing results are additionally shown as mutation frequencies (%). No material was available from the first time point at which the reported M184I was detected. NRTI = nucleoside reverse transcriptase inhibitor; NNRTI = non-nucleoside reverse transcriptase inhibitor; PI = protease inhibitor; 3TC = lamivudine; ZDV = zidovudine; DDI = didanosine; TDF = tenofovir; EFV = efavirenz; NFV = nelfinavir; DRV = darunavir; TPV = tipranavir; RTV/r = ritonavir/boosted ritonavir



paediatric HIV care. The predominant mutation among HIV-sDRM was M184V (32%), but other mutations affecting other drugs and classes were also found at varying rates, including TAMs (M41L, L210W, T215FY, L219E in 4.5 to 9%; D67N in 14%; K70R in 4.5%), discriminatory and nonpolymorphic NRTI mutations L74V and V75M, mutations affecting PIs (M46I in 18%; V82A and I54V in 9%) and NNRTIs (K101E, K103N, V106M, G190E, P225H, M230L in 4.5%). Only one case of INSTI DRM was found, in line with the study period of up to 2014, an era before the widespread availability of INSTIs in Switzerland. Although antenatal screening for HIV infection is recommended in Switzerland, eight women were not known to be HIV-infected during pregnancy and delivery, and the three remaining women did not receive antiretroviral prophylaxis during pregnancy for unknown reasons. Revisiting the MoCHIV data from the last 20 years, there was only one registered case of vertical HIV transmission after 2009, which occurred in 2011 (see [MoCHIV on SHCS website, fig. 3](#)). Together with the reduced rate of MTCT, the rate of HIV-tDRM is expected to decrease in Switzerland, but this cannot be generalised to other settings, e.g. resource-limited countries.

HIV-sDRM emerged on average three years after birth and was associated with more frequent treatment changes ($p = 0.104$), lower CD4 T-cell nadir ($p < 0.05$) and higher peak plasma HIV loads ($p = 0.06$). In the one case of HIV-tDRM, HIV-sDRM were subsequently detected after 10 months, suggesting that transmitted drug resistance might facilitate much earlier HIV-sDRM emergence and virological failure, although further analyses are needed due to the low sample size. The first report on HIV-tDRM was published in 1994, describing a case of neonatal infection by a zidovudine-resistant HIV-1 strain [24]. In a systematic review published in 2014, HIV-tDRM data among treatment-naïve children were only available from 14 countries, but the overall number of cases analysed was low [25]. As expected, the type and prevalence of HIV-tDRM reflected the choice and availability of antiviral treatments in the respective countries. Historically, the low cost of nevirapine administered as a single dose to the mother during labour and to the child right after delivery allowed its widespread use and improved the prevention of MTCT in low resource settings. Unfortunately, rapid selection of resistant viral mutants was soon observed [26, 27]. In high-income countries, HIV-tDRM of 10% to 17% have been estimated [15]. A recent study investigated both selected and transmitted HIV drug resistance and indeed found a rate of 4.9% in the latter group [22]. However, the data originated from a small cross-sectional study of 19 families and did not compare HIV-tDRM and -sDRM. There are few data on the role of HIV-tDRM, even though they potentially have effects on virological failure and the emergence of HIV-sDRM when only limited options for new antiretroviral drug classes exist.

Our in-depth analysis of HIV-DRM by NGS was limited to three patients with sufficient sample materials. The three cases were representative of three different scenarios. In two cases, HIV-DRM at frequencies below 20% that were not detected by Sanger sequencing were detectable by deep sequencing. However, in one case the HIV-sDRM emerged as a majority variant after two years and was then detected

by Sanger sequencing. Thus, NGS could be helpful for the identification of DRM-minority variants and for monitoring them over time. In the absence of DRM-informed decisions about effective cART, at least two new antiretrovirals that have not been used previously to treat either mother or child should be considered for children with virological failure [28].

Thus, the results of this study indicate that in the reported group of children, HIV-tDRM were found in only one out of 22 HIV-infected children born to mothers not receiving ART before and receiving it rarely during pregnancy (six with cART). In contrast, the frequency of HIV-sDRM was high and occurred with three to five years, despite the fact that half the children were treatment naïve.

This study has several limitations. Firstly, the number of available mother-child pairs was small, limiting the statistical analysis to the descriptive level, even though a long follow-up of 15 years was available. Secondly, this is a retrospective evaluation which included patients from a range of different eras of HIV diagnosis and management from the years 1989–2009, including the pre-ART and pre-cART eras. Thirdly, apparently treatment-naïve women appeared overrepresented among the 22 MTCT pairs, but this result emerged only because of this detailed analysis. However, we remain cautious by not directly attributing a low pre-test risk of HIV-tDRM to a patient history of no prior ART exposure. In some cases, there was no specific ART history available, whereas in others, this information may not have been reliable, given the stigma in patients with a difficult psycho-social history, including a migration background. Moreover, the transmission of drug-resistant viruses remains a possibility, especially in the early era of ART when insufficient suppression was usual. In this respect, it is notable that 60% of the MTCT pairs had the B subtype (fig. 2), but the 68% were of non-white ethnicity. Fourthly, HIV load and resistance testing could not be performed at all time points of interest, even though a significant effort was made by processing 109 samples of plasma and PBMC in addition to the HIV load data and resistance data that had been obtained at the time of the historic follow-up in the routine clinic. As the blood samples from these new-borns and infants were limited in size, additional next generation deep sequencing could not be performed except in 10 samples obtained from three cases. Nevertheless, the case studies were informative in principle, demonstrating that either no additional DRM were detectable (scenario 1), that DRM were detectable as minority species, but did not emerge as majority species (scenario 2), or that DRM were detectable as minority species and emerged as majority species during follow-up (scenario 3). Thus, HIV genotyping and resistance testing may provide relevant guidance for treatment, but access and adherence to appropriately dosed antiretrovirals, including new, not previously prescribed drug classes such as integrase inhibitors and combinations, remains an important consideration throughout paediatric HIV care.

Conclusions

While acknowledging these limitations, we conclude that HIV-tDRM were low in this Swiss study of mother-to child HIV transmission, making it a minor yet today preventable complication of HIV care [29, 30]. However, HIV-sDRM

remain an important issue in paediatric HIV care, underpinned by the fact that all options were available for the treatment-naïve children in our study. Indeed, similar to the paediatric care of other chronic medical conditions [31] such diabetes mellitus [32] and transplantation [33], non-adherence in paediatric HIV care remains challenging despite more potent drugs, lower pill counts and fewer side effects in the current era of cART [14].

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Potential competing interests

No potential conflict of interest relevant to this article was reported.

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